

Cytotoxicity of Different Degrees of De-Acetylated Chitosan on 3T3- and Two Human Tooth Fibroblast Cell-Lines

Research Article

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Abstract

Nowadays chitosan becomes more and more important as a drug carrier. In this study the influence of different degrees of de-acetylation of chitosan on the cytotoxicity were investigated.

Materials and Methods: Chitosan (87%, 70% and 40% de-acetylated) was analysed for their cytotoxic effect on mouse Balb/c 3T3 fibroblast cells as well as two different human tooth pulp fibroblast cell-lines. The cell survival rate was determined over a 24 hour period according to the standard MTT assay.

Results: The Univariate ANOVA test showed significant differences in the cell survival rates ($p < 0.01$) amongst 87% and 70% and 40% de-acetylated chitosan products for the 3T3 as well as the two pulp fibroblast cell-lines. Significant differences were also found between both tooth cell-lines and the 3T3 cell-line at all 3 different de-acetylated chitosan levels. The 3T3 cell line was mostly affected at 40% de-acetylation. The difference between both human cell-lines was not significant.

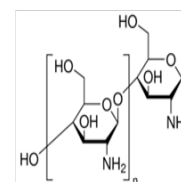
Conclusion: The lower the degree of chitosan de-acetylation the less the cell survival rate while an 87% de-acetylation degree improved the cell survival rate. Different cell lines react differently towards different degrees of de-acetylation. The relative survival rates of different cell lines changed at different degrees of de-acetylation.

Keywords: Chitosan; Degrees De-Acetylated; Cytotoxicity; 3T3; Tooth Fibroblasts.

Introduction

Due to the increasing applications of nanoparticles in many fields' becomes more and more under investigation. Especially, a growing concern is the possible adverse effects of exposure to nanoparticles. Therefore, cytotoxicity studies became more and more important. Of the most important factors to consider during such cytotoxicity studies are the cell type used, and in the case of chitosan, the degree of de-acetylation (DDA). Preferably the cell type under investigation should be the chitosan target cell.

Chitosan [1] is produced commercially by de-acetylation of chitin and is a natural cationic polysaccharide composed of randomly distributed β -1, 4-linked D-glucosamine and N-acetyl α -glucosamine.



This cationic nature is something special as most polysaccharides are neutral or negatively charged in an acidic environment [2]. The cationic nature allows it to form electrostatic complexes or multi-layer structures with other negatively charged molecules. Chitosan is non-toxic, biocompatible, bio-degradable and has mucous-adhesive properties and as a result became widely used in the pharmaceutical field as a carrier system for drugs, hormones, proteins, enzymes and genes [3-6]. Chitosan can be successfully used as a drug carrier because it will solubilize and degrade in an acidic environment with the resultant release of the drug [7]. However

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Citation: SR Grobler, A Olivier, HW Kruijse, VT Perchyonok (2016) Cytotoxicity of Different Degrees of De-Acetylated Chitosan on 3T3- and Two Human Tooth Fibroblast Cell-Lines. *Int J Dentistry Oral Sci.* 3(10), 337-339. doi: <http://dx.doi.org/10.19070/2377-8075-1600068>

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the low solubility in alkaline and neutral medium has a negative influence on the use. Chitosan is hypoallergenic and has natural antibacterial properties, which further support its use in the army as field bandages [8]. Furthermore, antioxidant-chitosan hydrogels (that of resveratrol, propolis and β -carotene) were found to significantly improve the bond strength to dentine with or without phosphoric acid pre-treatment [9] as many other hydrogels do [10].

Materials and Methods

Chitosan (Aldrich, Australia) were used as received. The degree of de-acetylation of typical commercial chitosan used in this study was 87% and the molecular weight 2500 kDa. The isoelectric point was 4.0–5.0.

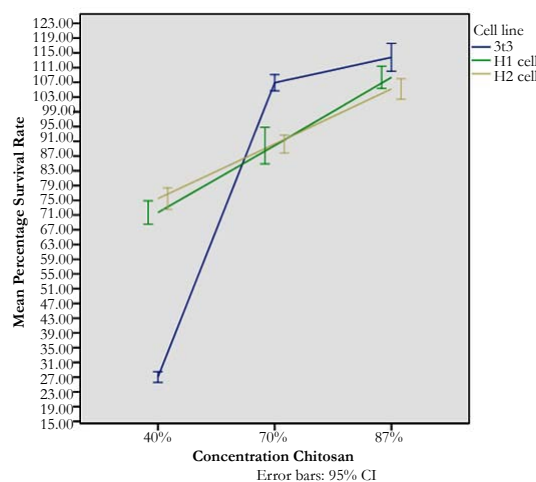
For the other de-acetylated products: Chitin was de-acetylated to ~70% (mw 45 kDa) and ~40% (mw 15 kDa) by the use of 45% sodium hydroxide solution in a 1:10 (W/V) ratio in a reaction vessel following Zhang’s method [11] and the reaction was under nitrogen atmosphere [12]. A Balb/c 3T3 mouse fibroblast cell line (The National Repository for Biological Materials, Sandringam) was maintained and cultured under standard conditions [13]. The two human dental pulp fibroblast cultures were established as previously described [14-16]. Briefly freshly extracted impacted 3rd molars were collected in Dubecco’s Modified Eagels Medium, the middle third of the pulp cultured and the cells between the 3rd and 5th passage used.

The cytotoxicity study was done as previously reported [13-16]. Briefly, the cells (mouse 3T3 Balb/c fibroblast, human tooth fibroblast 1 or human tooth fibroblast 2) were first grown to near confluency, diluted to a final cell suspension containing approximately 3×10^5 cells/ml and plated out in sets of 96 well plates. Chitosan 87% de-acetylated, 70% de-acetylated or 40% de-acetylated were then added to the growth medium at a concentration of 1 mg/ml. Two hundred μ l of each group was added to 20 wells in the 96 well plates. Medium without any gels was used as controls. After 24 hours the well-known MTT colorimetric assay was used to evaluate the cell survival rate. Absorbance was measured at wavelength 540 nm on a spectrophotometer to determine the number of viable cells. Three replicates were done in each group.

Results

The Figure shows the cell survival rates with 95% confidence

level error bars of the mouse Balb/c 3T3 fibroblast cell line, the human1 tooth pulp fibroblast cell-line and human2 tooth pulp fibroblast cell-line at 87% de-acetylated chitosan, 70% de-acetylated chitosan and 40% de-acetylated chitosan.



The results of a Univariate ANOVA showed significant main effects and an interaction effect suggesting an effect of increase in the concentration of chitosan.

Cell-line: $F_{(2,438)} = 23.71; p < 0.001$
 Concentration: $F_{(2,4378)} = 1120.8; p < 0.001$
 Cell-Line*Concentration: $F_{(4,438)} = 233.1; p < 0.001$

Significant differences were found between the two human cell-lines and the 3T3 cell-line at all 3 different de-acetylated chitosan levels. The difference between both human cell-lines is not significant at all 3 different de-acetylated chitosan levels (see Table below).

Discussion

We determined the survival rates at a growth medium concentration of 1mg/ml which is reported to be in line with other research and which may be considered as on the upper recommended level [16] of concentration. The three cell lines tested consisted of a standard cell line bought from the National Repository for Biological Materials and two self-established human tooth pulp fibroblast cell lines (H1 cell line and H2 cell line) from two different people. To our knowledge until now different degrees of de-acetylation towards the mentioned cell lines were not reported.

Cell-line	De-acetylation concentration	Mean	Std. Error	95% Confidence Interval	
				Lower Bound	Upper Bound
3t3	40%	27.028	1.406	24.264	29.791
	70%	106.894	1.179	104.577	109.212
	87%	115.399	1.456	112.538	118.26
H1cell	40%	72.370	1.422	69.574	75.165
	70%	89.883	1.473	86.987	92.779
	87%	107.714	1.198	105.360	110.069
H2cell	40%	74.807	1.208	72.433	77.181
	70%	90.260	1.362	87.584	92.936
	87%	103.127	1.492	100.196	106.059

Different degrees of de-acetylation (DDA) of chitosan is normally obtained by the treatment of chitin with alkali and a higher DDA is obtained by increasing the time and temperature, while the molecular weight (mw) of chitosan is dependent on the initial source of material which could be crab, fungi, shrimp etc. This mw normally decreases in the process to obtain a higher DDA [12]. In this study an 87% product with a mw of 2500 kDa was used as received, while the 70% (mw 145kDa) and 40% (mw 15kDa) were prepared. We did not investigate the effect of the molecular weight of the different chitosan's on the cytotoxicity.

It was reported that the degree of de-acetylation (DDA) and the molecular weight are important parameters which determines many physiochemical and biological properties of chitosans [12]. Nunthanid [17] reported that chitosans with high mw have higher moisture adsorption and tensile strength than those with the same DDA but lower mw. Hidaka [18] reported that a 94% DDA chitosan membranes gave minimal film degradation, mild inflammatory reaction and minimal osteogenesis while between 65 to 80% gave the opposite response. This effect is more or less in line to our survival rate results for all three different cell lines (Figure) that the higher the degree of de-acetylation the more the survival rate and that 87% de-acetylation actually improved the cell survival rate to above 100%.

However, Hamilton [19] reported no relationship between the DDA of chitosan, mw or growth of cells. While a similar report by the growth of keratinocytes was noted [20]. From the above it is concluded that there is controversial statements as far as the effect of molecular weights, in general, are concerned. In our study, the mw of the three chitosan products differed from 2500 kDa through 145 kDa to 15 kDa but this effect was not investigated.

In this study (Figure) we found that a 40% de-acetylation chitosan affects the 3T3 cell line significantly more (only about 30% cell survival was found) than both the two pulp cell lines. However, the other way round was found both at 70% and 87% de-acetylation where the pulp cell lines were significantly more affected (Figure). This showed that the relative survival rates of different cell lines can also change at different degrees of de-acetylation. The above finding is in line where it was reported that different cell lines have different levels of cytotoxicity towards a dentine bonding agent [14, 15]. In one study where 3 different human pulp fibroblast cell lines were studied it was found that one of these pulp cell lines were less cytotoxic than the 3T3 cell line but the other 2 were more sensitive.

Conclusion

The lower the degree of chitosan de-acetylation the less the cell survival rate while an 87% de-acetylation degree improved the cell survival rate. Different cell lines react differently towards different degrees of de-acetylation. The relative survival rates of different cell lines changed at different degrees of de-acetylation.

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