Hemorrhagic Centrolobar Necrosis and Cytoplasmic Vacuolation of the Hepatocytes in Syzygium Guineense Chronic Treated Mice

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Abstract

A World Health Organization survey indicated that about 70-80% of the world population rely on non-conventional medicine mainly of plant sources in their primary healthcare. In Ethiopia over 85% of the population relies on traditional medicine for the fight against various diseases. Syzygium guineense is one of such medicinal plants in use. It is traditionally used as a remedy for diarrhea, stomach pain and intestinal cramps. The toxic effects of these plants are often not put into considerations when they are being used. In this study we attempted to investigate the toxic effect of Syzygium guineense. On the histopathology of the liver of mice we choose the Liver because it is the major metabolic organ. Chronic administration of the aqueous of the extract of Syzygium guineense was found to be toxic to the hepatocytes as the micrograph showed increase vaculations, and shrinkage of the hepatocytes. There was also a general decrease in body weight and enlarged liver.

Keywords: Hepatocyte; Toxic; Syzygium; Guineense; Acqueous.

Abbreviations: EHNRI: Ethiopian Health and Nutrition Research Institute; ANOVA: Analysis of Variance.

Introduction

Plants that are commonly used in traditional medicine are assumed to be safe and their safety is based on their long usage in the treatment of diseases according to knowledge accumulated over centuries. However, recent studies have showed that many plants used as food or in traditional medicine are potentially toxic, mutagenic and carcinogenic [1, 2]. A World Health Organization survey indicated that about 70-80% of the world population rely on non-conventional medicine mainly of plant sources in their primary healthcare [3]. In Ethiopia over 85% of the population relies on traditional medicine for the fight against various diseases [4]. Plant remedies still remain the most important and sometimes the only source of therapeutics for nearly 80% of the population, in Ethiopia and the Sub Saharan Africans [5].

In Ethiopia, there are traditional medicines that have been used by many folk medicine practitioners without taking into consideration their side effects on different parts of the body [6]. Syzygium guineense is one of such medicinal plants in use.

S. guineense grows in most parts of Ethiopia at altitudes of 1200-2500m [7] and is traditionally used as a remedy for diarrhea, stomach pain and intestinal cramps [6].

Both the hydroalcoholic and aqueous extracts of the leaves of S. guineense was found to be effective at the dose of 200mg/kg in controlling castor oil induced diarrhea in mice by oral administration [8]. Same study also demonstrated that the leaf and stem bark extracts had spasmylytic effect in Guinea pig ileum.

Phytochemical investigation of S. guineense showed the presence of tannins, phytosteroids, flavonoids and saponins in some of the crude extracts and fractionates [8]. Eugenin, an active ingredient found at buds of Syzigium Spp; had antiviral effect against Herpes simplex [6, 9]. According to [10] Syzgyium guineense had potent...
antibacterial effect against diarrhea causing bacteria. Methanolic extract of *S. guineense* inhibited intrinsic contraction in isolated ileum tissue of* rabbit [11]. *Eugenia guineense* revealed a significant increase in the plasma bilirubin of alkaloidal fraction treated rats, which is suggestive of a possible damage to the liver occasioned by the treatment with the *E. guineense* stem-bark total alkaloidal fraction [12]. The aim of the present study was to investigate the effect of the extract of *S. guineense* on the histopathology of the liver of mice.

**Materials and Methods**

**Experimental design and Laboratory**

It is an experimental case control type of research. The animal laboratory works; histological tissue processing was done in the histology laboratory of anatomy department of Mekelle University Ethiopia. Light microscopy and micrographs were taken in the histology laboratory of Anatomy Department, Kogi State University Anyigba Nigeria.

**Collection of Plant Materials**

Fresh leaves of *S.guineense* were collected from Woliso area about 116 km south west of Addis Ababa, Ethiopia, during the month of February, 2015. Plant sample was identified and deposited in the National Herbarium of the Department of Biology, AAU with a specimen number of 001, 002, and 003.

**Processing of the Plant Materials**

Fresh leaves of *S.guineense* were cleaned with tap water from the extraneous materials, dried at room temperature, and ground to powder in the Phytochemistry Laboratory of the Department of Drug research, Ethiopian Health and Nutrition Research Institute, EHNRI. Extraction of the aqueous extract of the ground fresh leaves was done by maceration of 600 g in 2000 ml of distilled water. The aqueous extract was intermittently agitated for 24 hours by Orbital shaker (DS-500 model 0.75” 1.9cm Cat. No. 444.7018) at room temperature. After 24 hrs, the extract was filtered with gauze and lyophilized to give a powdered material yield of the extract, which was 9.5%. The extract was then kept in desiccators at room temperature till use.

**Experimental Animals**

The research was conducted following the approval of the proposal by the Faculty Research and Publication Committee. Male and female animals of Swiss albino mice (25-30 g) were provided from the animal department of EHNRI. The mice were acclimatized for 5 days to the environment of the animal house in the department of drug research, EHNRI. Animals were fed standard commercial diet and water *ad libitum*. They were maintained at standard conditions of temperature (21 ± 2°C), relative humidity of 65 ± 0.5% and 12 hours light/night cycles till the end of the experiment. The mice were randomly divided into four groups with each group consisting of 10 mice (5 male and 5 female) and separate cages were used for the sexes. Group I served as a control and received the vehicle, i.e., distilled water. Groups II, III and IV were given 200, 400 and 600 mg/kg bw of the extract for each group, respectively, once a day at 24 hour intervals, for six weeks. Administration of the extract was done with intragastric tube, and the volume administered in all cases was 0.5 ml/mouse. The no-effect dose was that inhibited diarrhea in the * in vitro * study carried out for the effect of crude leaf extract of *S. guineense* on castor-oil induced diarrhea [8].

At the beginning and end of six weeks, all the mice were weighed as described by [13]. Body weight taken on the initial date of treatment was considered the initial weight and the weight taken on the last day of administration before anesthesia and dissection; was considered the final weight.

**The LD₅₀ and Tissue Morphology**

A batch of 28 mice (27-30g) of both sexes were divided randomly into two groups of each containing 14 mice (7 male and 7 female), one of which serving as a control. The oral LD₅₀ (median lethal dose) of the aqueous extract was 14.1g/kg bw [15]. The extract was given orally after overnight fasting. The control group was given distilled water orally. Soon, after the death of the animal, dissection was carried out. The tissue samples were collected from the liver and immersed immediately in a fixative (10% buffered neutral formalin solution).

**Animal Dissection and Tissue Sample Collection**

Animals were anesthetize and sacrifice. The whole liver was exposed and carefully removed and blotted on filter paper and weighed quickly on a semi-microbalance sensitive to 0.001mg (SciENTECH Boulder Co., USA). Part of the median lobe of the liver were cut in coronal and transverse sections respectively and placed immediately in fixative (10% buffered neutral formalin solution) for tissue processing. Similarly, the liver of mice found dead was also dissected immediately, trimmed and placed using the same methodology.

**Tissue Processing**

The tissues were processes following routine tissue processing techniques for hematoxyline and eosine. This was done in the histology laboratory of Anatomy department Mekelle University.

**Light Microscopy**

Sections of the liver and kidney were examined under the compound light microscope with different objectives and photomicrographs of selected samples of the liver and kidney tissue were then taken by Fuji iso 200 film using a MC 80 DX Microscopic Camera (Carl Zeiss Jena Gmbh, Germany) and Zeiss Aploscope microscope fitted with a digital camera (Nicon Coolpix 5000). A magnification of ×40 was used to examine changes in the liver tissue. Following the evaluation of control animals, the liver of the remaining groups were evaluated blind to treatment.

**Statistical Analysis**

All the values in the test are expressed as mean ± SEM. Statistical differences between the means of various groups were evaluated by one-way analysis of variance (ANOVA) using the SPSS version 11.0 programmer followed by student’s *t*-test. P-values less than 0.05 were considered to be significant.
Results

The results observed from this study are tabulated below and the micrographs are also presented below.

Light microscopic examination of liver tissue of the control mice showed characteristic features showing normal portal triad, interlobular bile duct, radiating hepatic cells and hepatic sinusoids lined by endothelial cells (Figure 1A).

Mice treated at a dose of 200 and 400 mg/kg bw showed no histopathological changes as compared to the control (Figure 1, B and C). However, tissue morphology of mice treated with 600 mg/kg bw of the extract showed hemorrhagic necrosis, and cytoplasmic vacuolations (Figure 1D). Similarly, congestion of liver sinusoids was observed in mice treated with the LD₅₀ of the extract (Figure 2).

Discussion

In this study, significant general body weight gain of mice was observed in mice treated with 200, and 600mg/kg bw as compared to the control group as shown in Table 1. The weight gain difference between the treated and the control groups suggest the presence of toxic constituents in the extract that impaired the growth. This is in agreement with the findings of [14] that rats on 10% *Acacia abyssinica* diet which showed decrease in body weight gains and development of hepatop-nephropathy indicating that the plant contains toxic constituents that impaired growth and reduced their rate of excretion by the injured liver.

The increase in liver weight with 200 and 400mg/Kg extract is due to the toxic effect of the active component(s) present in the extract leading to abnormal histology of the liver. The increased in liver weight in the present study as shown in Table 2, is consistent

<table>
<thead>
<tr>
<th>Group</th>
<th>Treatment (mg/kg bw)</th>
<th>Initial body weight (g)</th>
<th>Final body weight (g)</th>
<th>Weight difference (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Control</td>
<td>26.68 ± 1.69</td>
<td>32.95 ± 2.95</td>
<td>4.27 ± 3.29</td>
</tr>
<tr>
<td>II</td>
<td>200</td>
<td>28.27 ± 1.6</td>
<td>29.3 ± 3.7*</td>
<td>0.09 ± 3.26</td>
</tr>
<tr>
<td>III</td>
<td>400</td>
<td>27.52 ± 1.23</td>
<td>29.08 ± 2.16</td>
<td>1.47 ± 2.99</td>
</tr>
<tr>
<td>IV</td>
<td>600</td>
<td>26.29 ± 2.09</td>
<td>27.31 ± 2.67*</td>
<td>0.52 ± 3.78</td>
</tr>
</tbody>
</table>

Values are mean ± SEM. N = 10/group, *N = 9/group.

<table>
<thead>
<tr>
<th>Group</th>
<th>Treatment (mg/kg bw)</th>
<th>Relative organ weight (gms/100g)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Liver</td>
</tr>
<tr>
<td>I</td>
<td>Control</td>
<td>3.77 ± 0.26</td>
</tr>
<tr>
<td>II</td>
<td>200</td>
<td>4.68 ± 0.59*</td>
</tr>
<tr>
<td>III</td>
<td>400</td>
<td>4.31 ± 0.37</td>
</tr>
<tr>
<td>IV</td>
<td>600</td>
<td>4.13 ± 4.21*</td>
</tr>
</tbody>
</table>

Values are mean ± SEM. N = 10/group, *N = 9/group.

Figure 1. Photomicrographs of mice liver (*H & E* ×4216).

Effects of the extract on tissue morphology of the liver H = Hepatocyte, S = Sinusoid, CV = Central venule, B = Hemorrhage, V = Cytoplasmic Vacuolations, E = Endothelium, N = Necrosis.
Effects of the extract at LD50 dose on tissue morphology of the liver H = Hepatocyte, S = Sinusoid, CV = Central venule, B = Hemorrhage, V = Cytoplasmic Vacuolations, E = Endothelium, N = Necrosis

with the ones observed in the previous studies done with other plants [15-17].

Chronic administration of 600mg/kg S. guineense aqueous leaf extract in mice revealed centrilobular necrosis of the hepatocytes of the liver, which is in agreement with that of [18], who reported that massive hepatocyte necrosis was observed in the liver of the rats treated with aqueous extract of Securidaca longepedunculata and described that this abnormal histology of the liver is characterized by focal hepatocellular necrosis similar to the pattern of damage induced by overdose of paracetamol [19].

Explained hepatotoxic drugs and chemicals may act through their metabolites which could typically cause centrilobular necrosis. They demonstrated an autopsy specimen of acetaminophen that disclosed a prominent hemorrhagic necrosis of the centrilobular zones of all liver lobules. A study by [20] reported that mice treated with Senna podocarpus extract revealed congested central veins and sinusoids of the liver.

Conclusion

The aim of this study was to investigate the effect of the extract of Syzygium guineense on the histopathology of the liver of mice. From the result obtained here and in comparison to other studies we conclude that chronic usage of S. guineense is toxic to the hepatocytes, and will result to decrease body weight and enlarged liver.

References


