

**Effect of Some Antihypertensive Drugs on Alkaline Phosphatase and DNA of Mice**

**Research Article**

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**Abstract**

The dislipidemias are a risk factor well recognized of the cardiovascular diseases and constitute a problem of public health. A descriptive study in 150 patient elders of 30 years with diagnosis of Izuemic Cardiopathyes accomplished itself for the sake of identifying dislipidemias in patients of high cardiovascular risk that they helped the high-technology General Medical Center state James Mariño Aragua, at the Republic Bolivariana of Venezuela, that you constituted the sign of study from October 2011 to October 2012. They used quantitative and qualitative variables like weight, age, sex, pathological personal background, risk factors cardiovascular associates, seric levels of total cholesterol, triglycerides, HDL cholesterol, LDL cholesterol VLDL cholesterol. 63 percent of patients with dislipidemias were detected, being hypercholesterolemia the more alteration frequently found. The ages understood between 41 and 60 years evidenced the bigger frequency.

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**Introduction**

Cardiovascular diseases are one of the leading causes of death for people in developed countries world-wide. Hypertension and osteoporosis are frequent diseases among elders. Both are induced by interaction of many genetic and environmental factors (Zhang et al., 2007). Many metabolic and physiological changes occur in hypertensive individuals and some of these processes stem from the associated increased oxidative stress ( Vasdev, 2006). The depression in antioxidant enzymes and increase in oxidants in the hypertensive state have been reported to increase the production of reactive oxygen species (Lassegue and Griendling, 2004 ; Manning et al., 2005 ). The reactive oxygen species/free radicals resulting from the oxidant-antioxidant imbalance tend to accumulate and are known to cause oxidative damage to the cellular macromolecules including the genetic material (Rao, 2009). The genetic instability can include mutations, chromosomal aberrations and unscheduled DNA synthesis (Prerara and Bapat, 2007 ; Shimizu et al., 2008). Amlodipine is a derivative of dihydropyridine and is one of calcium channel blockers. Its principal action is to inhibit

calcium entry through voltage-gated transmembrane L-type channels, thus decreasing intercellular calcium concentration and inducing smooth muscle relaxation (Liu et al., 2007; Devabhaktuni and Bangalore, 2009; Liu et al., 2011). Atenolol is a beta 1-selective drug that binds to the beta-receptors and prevents the stimulation by catecholamine resulting in lowering of heart rate and of the systemic blood pressure (Wadworth et al., 1991). Captopril is an angiotensin-converting enzyme (ACE) inhibitor used for the treatment of hypertension and some types of congestive heart failure. Captopril was the first ACE inhibitor developed and was considered a break through both because of its novel mechanism of action and also because of the revolutionary development process (Akif et al., 2010).

Alkaline phosphatase (ALP) is an enzyme that catalyzes the hydrolysis of organic pyrophosphate and inhibitor of vascular calcification (Harmey et al., 2004). Although ALP is expressed in a variety of tissues, its concentrations are highest in bone, liver and kidney (Schoppet and Shanahan, 2008). Accordingly, serum levels of ALP are used in clinical practice as a marker of hepatic , peripheral vascular and bony diseases (Regidor et al., 2008). No published data have been found in the relevant literature on the toxicity of anti-hypertensive drugs on the DNA of normal mice and thereby the present study was undertaken to determine the effect of chronic administration of recommended ; half recommended and one and half recommended doses of the antihypertensive drugs amlodipine, atenolol and captopril on the genomic DNA and on the activity of alkaline phosphatase (ALP) in the liver, spleen and kidney of Swiss albino mice.

**Materials and Methods**

**Chemicals**

Atenolol was purchased from Kahira Pharmaceutical. and Chemical Industries Company (Egypt). Amlodipine was purchased from Pfizer (Egypt) and captopril was purchased from Bristol My-

ers Squibb (Egypt). All other chemicals were of analytical grade.

### Experimental animals

All experiments were performed using adult female Swiss albino mice, with an average body weight of 25 g purchased from Theodore Bilharz Research Institute, Giza, Egypt. The mice were housed in steel mesh cage and provided with commercial standard diet and tap water ad libitum.

A total of 150 Swiss albino mice were divided into 10 groups, 15 mice each, according to the following scheme: group 1, control (untreated) mice; groups 2-4, mice received captopril in doses of 1.3 mg/kg b.w. (low dose; half the recommended dose), 2.6 mg/kg b.w. (the recommended dose) and 3.9 mg/kg b.w. (high dose; one and half times the recommended dose) respectively every other day; groups 5-7, mice received amlodipine in doses of 0.26 mg/kg b.w. (low dose; half the recommended dose), 0.52 mg/kg b.w. (the recommended dose) and 0.78 mg/kg b.w. (high dose; one and half times the recommended dose) respectively every other day and groups 8-10, mice received atenolol in doses of 5.2 mg/kg b.w. (low dose; half the recommended dose), 10.4 mg/kg b.w. (the recommended dose) and 15.6 mg/kg b.w. (high dose; one and half times the recommended dose) respectively every other day. Each of the nine groups of mice receiving the drugs were classified into subgroups, 5 animals each. The animals of the subgroups were sacrificed at 1, 3, 6 months of starting administration of the drugs while the animals of the control group were all killed at the end of the experimental period. Liver, spleen and kidney were immediately excised, washed in ice-cold saline, blotted dry and weighed for measuring various biochemical parameters.

### Preparation of homogenates

An accurately weighed piece of each of liver, spleen and kidney tissues was homogenized in ice-cold 0.9 % saline using a Teflon pestle connected to a homogenizer motor. The homogenates were adjusted at a concentration of 5 % (w/v). The homogenates were centrifuged at 5000 rpm for 30 minutes at 4 °C to remove cell debris and nuclei. The resulting supernatant was used for biochemical analysis.

### Biochemical Analysis

Alkaline phosphatase activity was estimated by the method of Ali et al. (2006) using a commercial available assay kit (AAT Bioquest®, Inc., Egypt). Protein in the homogenates was determined by the method of Lowry et al. (1951).

### Genomic DNA extraction and RAPD analysis

Total DNA from mice tissues was extracted and purified according to the Wizard® Genomic DNA Purification Kit supplied by Promega Corporation (USA). Among several primers used for screening and RAPD-PCR analyses of the studied samples only one primer OPE-13: 5'- CCCGATTCGG-3' was chosen. Genomic DNA was analysed with random amplified polymorphic DNA polymerase chain reaction (RAPD-PCR) technique according to the method described by Williams et al. (1990). The PCR amplifications were performed in 25 µl of the reaction mixture containing 1 µl of template DNA, 3.5 µl primer and pure taq Ready-To-Go PCR beads. The PCR conditions were as follows: denaturing at 94 °C for 1.5 min, annealing at 37 °C for 1.5 min and extension at 72 °C for 3 min. The products were mixed

with the loading buffer and were loaded in 2% agarose gels and electrophoresed at 100 V for 1h. The gels were stained with ethidium bromide and visualized under UV light.

### Statistical analysis

The results are expressed as means ± SD. Statistical analysis was performed according to the method of Murray (1982). Data were analyzed using unpaired Student's t-test. P values of < 0.05 were considered to be statistically significant.

### Results

Table 1 illustrates the alkaline phosphatase (ALP) activities in mice of groups. After administration of captopril for one, three and six months in doses of 1.3, 2.6, 3.6 mg/kg b.w., highly significant elevations in the activities of ALP in the liver, spleen and kidney homogenates were found compared to its activities in the homogenates of the control group. Similarly, a highly significant increases in the activities of ALP in the liver and spleen homogenates of mice were observed after administration of 5.2, 10.4, 15.6 mg/kg b.w. atenolol for one, three and six months. On the other hand, a significant decrease was observed in ALP activity in the kidney homogenates after administration of 5.2 mg/kg b.w. atenolol for one month while no significant changes were found after three and six months from the administration of atenolol. Moreover, a significant decrease in the kidney ALP activity on the administration of 10.4 and 15.6 mg/kg b.w. of atenolol for six months was found compared to the control activity. The administration of 0.26, 0.52 and 0.78 mg/kg b.w. of amlodipine for one, three and six months caused highly significant increases in the activities of ALP in the liver, spleen and kidney homogenates compared to the corresponding activities of the controls.

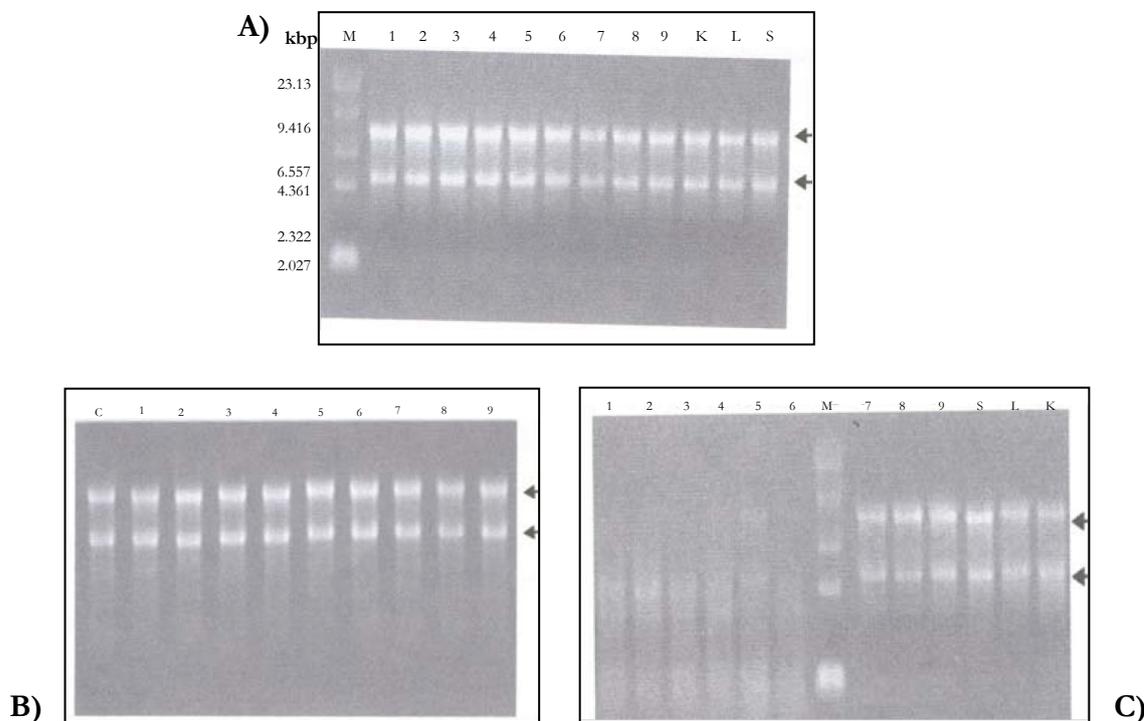
The results of RAPD-PCR for DNA samples isolated from liver, kidney and spleen of control mice and mice treated for 1, 3 and 6 months with various doses of captopril, atenolol or amlodipine are shown in Fig 1A, B, C. The amplification of products obtained by this technique showed the presence of numerous bands between 2 and 23 kbp (Fig 1C). The primer gave a recognizable profile of bands for amlodipine treated samples (Fig 1C) for 6 months compared to atenolol, captopril (Fig 1 A & B) and control samples (Fig 1 A & C). Overall, the treatment by amlodipine caused more changes in RAPD-PCR patterns than treatment by either captopril or atenolol especially in liver, spleen and kidney.

### Discussion

The hypertension increases the risk for stroke and coronary heart disease and is a main contributor to pre-mature death (Yadav et al., 2008). Toxic effects of drugs may be functional, biochemical, structural and specific. Such effects are evaluated by assessment of oxidant-antioxidant parameters in tissues and severity of DNA damage (Polat et al., 2010). The pathology of essential hypertension may involve one or more abnormalities in cardiovascular homeostatic mechanisms including endothelial dysfunction (Beavers et al., 2001). The endothelial dysfunction is related to a defect in vasodilator nitric oxide (NO) synthesis. There may be an elevation of reactive oxygen species like superoxide anion, which inactivate the NO. Normally, the endogenous antioxidant mechanism involving enzymes like catalase and superoxide dismutase in addition to reduced glutathione scavenge and regulate the superoxide formed. This protective mechanism may be defective in essential hypertension cases (Touyz, 2004). In the

Groups	Alkaline Phosphatase (IU)								
	1m			3m			6m		
	L	S	K	L	S	K	L	S	K
Group 1 (Control)	14.7 ± 1.81	3.88 ± 0.70	3.36 ± 0.947	14.7 ± 1.81	3.88 ± 0.70	3.36 ± 0.947	14.7 ± 1.81	3.88 ± 0.70	3.36 ± 0.947
Group 2 (Treated with Cap-topril 1.3mg/kg)	18.6 ± 0.55 **	11.59 ± 2.3 **	16.08 ± 0.55 **	17.13 ± 2.82 **	11.88 ± 0.11**	11.55 ± 1.06 **	54.93 ± 3.98**	8.6 ± 1.22 **	12.52 ± 1.18 **
Group 3 (Treated with Cap-topril 2.6mg/kg)	25.14 ± 0.75 **	15.96 ± 0.54 **	15.5 ± 1.98 **	32.32 ± 4.009 **	16.9 ± 1.34 **	19.29 ± 1.4 **	26.24 ± 1.046 **	19.33 ± 1.27 **	16.14 ± 1.605 **
Group 4 (Treated with Cap-topril 3.9mg/kg)	34.99 ± 1.65 **	31.39 ± 1.5 **	10.98 ± 0.92 **	61.55 ± 3.45 **	35.19 ± 2.21 **	16.76 ± 0.72 **	26.76 ± 1.18 **	6.4 ± 1.83 **	5.47 ± 1.025 **
Group 5 (Treated with Aten-olol 5.2mg/kg)	20.83 ± 0.48 **	8.84 ± 0.86 **	1.08 ± 0.176 *	43.9 ± 0.346 **	7.09 ± 0.72 **	3.98 ± 0.339 NS	17.58 ± 1.06 **	8.80 ± 0.78 **	3.17 ± 0.784 NS
Group 6 (Treated with Aten-olol 10.4mg/kg)	25.34 ± 1.83 **	12.66 ± 1.4 **	18.57 ± 1.59 **	70.76 ± 1.89 **	5.93 ± 0.28 **	14.414 ± 1.166 **	18.74 ± 0.40 **	26.45 ± 0.905 **	1.86 ± 0.21 **
Group 7 (Treated with Aten-olol 15.6mg/kg)	29.81 ± 0.94 **	14.08 ± 1.59 **	12.51 ± 1.27 **	53.37 ± 1.26 **	9.916 ± 1.011 **	13.62 ± 0.33 **	24.014 ± 0.59 **	30.5 ± 0.558 **	1.93 ± 0.006 **
Group 8 (Treated with Amlodipine 0.26mg/kg)	61.2 ± 0.593 **	10.25 ± 0.41 **	10.0 ± 4.02 **	51.9 ± 3.04 **	5.314 ± 0.021 **	18.5 ± 0.39 **	19.6 ± 1.73 **	10.97 ± 0.813 **	23.05 ± 0.629 **
Group 9 (Treated with Amlodipine 0.52mg/kg)	9.23 ± 0.72 **	25.3 ± 0.014 **	6.23 ± 1.407 **	16.6 ± 0.84 **	6.48 ± 0.148 **	7.96 ± 1.42 **	17.69 ± 1.54 **	6.39 ± 0.417 **	16.6 ± 1.32 **
Group 10 (Treated with Amlodipine 0.78mg/kg)	19.97 ± 8.06 **	9.48 ± 0.49 **	9.22 ± 2.33 **	18.6 ± 0.134 **	9.89 ± 1.25 **	9.54 ± 2.13 **	21.2 ± 0.226 **	19.7 ± 0.74 **	5.08 ± 0.95 **

**Figure 1: RAPD- PCR analysis of DNA samples isolated from livers (lanes 1-3); spleen (lanes 4-6) and kidney (lanes 7-9) of capotpril –treated (A) ; atenolol- treated (B) and amlodipine-treated mice (C) : Lane M :standard molecular weight marker . Lanes K, L, S in Fig. A and C: kidney, liver and spleen DNA of control mice. Lane C in Fig. B: liver DNA of control mice. Lanes 1, 4, 7 : samples after one month treatment ; Lanes 2, 5, 8: samples after 3 month treatment; Lanes 3, 6, 9: samples after 6 month treatment.**



present study the effects of treatment with the antihypertensive drugs amlodipine, atenolol and captopril on alkaline phosphatase (ALP) activity and genomic DNA was investigated in liver, spleen and kidney of mice. After chronic administration of captopril and amlodipine, the activity of ALP in liver, spleen and kidney homogenates were highly significantly increased compared to its level in the organs of mice. These findings are in accordance with those reported by Nishiya and Sugimoto, (2001), Gradosova et

al. (2011) and Moraes et al. (2011). Tonelli et al. (2009) reported that the higher level of ALP can be used as a marker for patients with an underlying inflammatory state, vascular calcification and as a liver function. Some investigators showed that the chronic administration of captopril affected the promotion of the secretion of ALP and its level depends on the dose of captopril (Liu et al. 2011). The present results demonstrated that chronic administration of atenolol exhibited significant increase in the activity of

ALP. These results are in good agreement with those of other investigators (Mota et al., 2008). The elevation of ALP activity in normal mice after administration of antihypertensive drugs may reflect a link between the abnormal mineral metabolism of kidney and excess cardiovascular mortality (Ali et al., 2006 & Blayney, et al., 2008).

The analysis of genomic alterations in liver, spleen and kidney of mice treated with atenolol and captopril using RAPD-PCR fingerprinting showed that these drugs did not produce significant changes in the RAPD-PCR fingerprints of the liver, spleen and kidney genomes. Both drugs produced nearly the same DNA banding patterns similar to that of control (non-treated) mice. However, amlodipine exerted an adverse affect on the DNA content of the liver, spleen and kidney. Apparently, amlodipine degraded the liver, spleen and kidney DNA and masked the sites for the annealing of the RAPD primers. These present findings are disagreeable with previous studies reporting that long term therapy of atenolol increased expression of fragile sites of DNA and caused chromosomal aberrations (Telez et al., 2000 ; Telez et al., 2010). On the other hand, the present results are in consistent with the results of other investigators (Kang et al., 1997 & Liu et al., 2006) who reported that atenolol and captopril did not show any significant antiproliferative effect on the cultured rat aortic smooth muscle cells through-out any of the studied doses. Many studies indicated that the calcium channel blocker amlodipine influences DNA and affect bone tissues resulting in decrease in bone resorption (Rejnmark et al., 2006 ; Ushijima et al., 2010 ; Subash et al., 2010). In these previous reports, accumulating data suggested that this calcium channel blocker (amlodipine) has anti-inflammatory-antioxidant and antiapoptotic properties beside its antihypertensive action (Yamagata et al., 2004 & Yoshii et al., 2006). To summarize, our results clearly indicated that the chronic administration of amlodipine affected the genomic DNA and the activity of ALP. The present results may be attributed to: [1] the balance between inorganic pyrophosphate and serum phosphate which regulates vascular calcification. [2] imbalance in oxidant and antioxidant levels due to hypertension which resulted in accumulation of DNA damage with time ( Khanna et al., 2008). In addition it is concluded that the studied antihypertensive drugs may induce inflammation by increasing the activity of ALP.

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